SNAP-25 Is a Target of Protein Kinase C Phosphorylation Critical to NMDA Receptor Trafficking

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Protein kinase C (PKC) enhances NMDA receptor (NMDAR)-mediated currents and promotes NMDAR delivery to the cell surface via SNARE-dependent exocytosis. Although the mechanisms of PKC potentiation are established, the molecular target of PKC is unclear. Here we show that synaptosomal-associated protein of 25 kDa (SNAP-25), a SNARE protein, is functionally relevant to PKC-dependent NMDAR insertion, and identify serine residue-187 as the molecular target of PKC phosphorylation. Constitutively active PKC delivered via the patch pipette potentiated NMDA (but not AMPA) whole-cell currents in hippocampal neurons. Expression of RNAi targeting SNAP-25 or mutant SNAP-25(S187A) and/or acute disruption of the SNARE complex by treatment with BoNT A, BoNT B or SNAP-25 C-terminal blocking peptide abolished NMDAR potentiation. A SNAP-25 peptide and function-blocking antibody suppressed PKC potentiation of NMDA EPSCs at mossy fiber-CA3 synapses. These findings identify SNAP-25 as the target of PKC phosphorylation critical to PKC-dependent incorporation of synaptic NMDARs and document a postsynaptic action of this major SNARE protein relevant to synaptic plasticity.

Introduction

NMDARs are glutamate-gated ion channels and play a pivotal role in the regulation of synaptic function in the brain. NMDARs are heteromeric assemblies of NR1, NR2, and NR3 subunits, which cotranslationally assemble in the endoplasmic reticulum (ER) to form functional channels with differing physiological and pharmacological properties (Wenthold et al., 2003; Pérez-Ótano and Ehlers, 2005; Lau and Zukin, 2007). NMDAR-mediated Ca2+ influx is essential for synaptogenesis, experience-dependent synaptic remodeling, and long-lasting changes in synaptic efficacy such as long-term potentiation (LTP) and depression (LTD), cellular processes widely believed to underlie learning and memory (Collingridge et al., 2004; Malenka and Bear, 2004). Recent studies indicate that NMDARs serve not only as a trigger of synaptic plasticity, but may also contribute to the expression of LTP and LTD (Lau and Zukin, 2007).

PKC plays a critical role in memory formation and storage (Wang et al., 2006), induction of LTP (Ben-Ari et al., 1992), and vesicular exocytosis (Morgan et al., 2005). PKC increases NMDAR channel opening rate in Xenopus oocytes and promotes NMDAR insertion at the cell surface via SNARE-dependent exocytosis (Lau and Zukin, 2007). Although NMDARs are a known functional target of PKC potentiation, the direct molecular target of PKC phosphorylation is unclear. Target PKC and PKA phosphorylation sites have been identified on the NR1, NR2A, and NR2B subunits (Leonard and Hell, 1997; Tingley et al., 1997). However, NMDARs assembled from mutant subunits lacking all known sites of PKC phosphorylation show marked PKC potentiation (Zheng et al., 1999; but see Liao et al., 2001), suggesting that the molecular target of PKC is a receptor-associated signaling and/or trafficking protein.

Emerging evidence indicates a role for the SNARE family of membrane fusion proteins in trafficking of postsynaptic glutamate receptors (Lledo et al., 1998; Lan et al., 2001; Lu et al., 2001; Washbourne et al., 2004). The synaptosomal-associated protein of 25 kDa (SNAP-25) is a member of the SNARE family and a known target of PKC phosphorylation (Shimazaki et al., 1996). SNAP-25 assembles with syntaxin-1 and vesicle-associated membrane protein-2 (VAMP2/synaptobrevin–2) via SNARE motifs to form the SNARE complex, the minimal machinery necessary for exocytosis of neurotransmitter vesicles at the presynaptic terminal (Jahn et al., 2003). SNAP-25, syntaxin-1, and VAMP2 localize...
to biochemical fractions of postsynaptic densities (PSDs), consistent with the possibility that they also participate in trafficking of postsynaptic receptors (Jordan et al., 2004; Cheng et al., 2006).

The present study was undertaken to identify the molecular target of PKC phosphorylation functionally relevant to PKC-induced insertion of NMDARs at the cell surface of neurons. SNAP-25 is implicated in vesicular fusion and/or docking at presynaptic terminals (Washbourne et al., 2002a) and PKC phosphorylation of SNAP-25 is critical to PKC-induced, SNARE-dependent insertion of NMDARs at synaptic sites, a mechanism relevant to synaptic plasticity.

Materials and Methods

DNA constructs. NR1,100 (NR1-4b) and NR2A cDNAs in pBluescript SK (−) vector were linearized at the 3′ end with BamHI and NotI, respectively, and used for in vitro transcription of mRNA. WT and Δ20 (dominant-negative) mutant constructs of SNAP-25 in pcDNA3 were gifts from Dr. Roger Y. Tsien (Howard Hughes Medical Institute, University of California, San Diego, CA). SNAP-25 constructs were linearized at the 3′ end with EcoRI. Capped mRNAs were synthesized as runoff transcripts from linearized plasmid cDNAs with T7 polymerase (mMessage mMACHINE transcription kit; Ambion; 2 h at 37°C).

Mutagenesis. The SNAP-25 protein sequence harbors 3 sites that conform to the PKC consensus phosphorylation motif (S/T-X-R/K): Ser28, Thr29 and Ser187. These residues in SNAP-25 were mutated into Ala or Cys by site-directed mutagenesis, singly or altogether using the QuikChange XL Site-Directed Mutagenesis Kit according to the manufacturer’s protocols (Stratagene). The mutagenic primer sequences for the S187A mutation were (5′ to 3′): GATCATGGAGAGGTGCAGGC-CACAAACACAGAATGTG and CAATCTCTGGTTTTTGCTGCTAGC and GCTTCTGCCATGTC; for S28AT29A were GGCTGAC-CTGACTGAC and TACAGGGACACACACAAAGCCG; for S187A and S28AT29A were GGCTGAC-CTGACTGAC and GCAGCATGCTGC and GCAGCAT-GGACGCGGCGGCCTCGAGCGATCAGCC.

Introduction of an XhoI site (underlined) in the S28AT29A primers allowed fast screening of the mutant by restriction digest.

Subcloning. For transfection into HEK-293 cells and neurons in culture, WT and mutant SNAP-25 cDNAs were subcloned into the pIRE2-EGFP vector (BD Biosciences Clontech), which allows production of a single bicistronic mRNA containing SNAP-25 and EGFP. SNAP-25 cDNAs were excised from pcDNA3 with BamHI (5′) and EcoRI (3′), and pIRE2-EGFP was cut with BglII (5′) and EcoRI (3′) enzymes and the 3′ end dephosphorylated by CIP. The fragments were ligated together overnight at 16°C using T4 ligase (Promega) or Quick Ligation kit (New England Biolabs) at a molar ratio of 1:3 (vector:insert). The final constructs and orientation of inserts were confirmed by direct sequencing.

Lentiviral-mediated SNAP-25 RNAi. Double-stranded RNA sequences targeting distinct regions of mouse SNAP-25 (engineered to mimic endogenous microRNA structure) were subcloned into a pcDNA6.2-GW/EmGFP-miR vector (pH pimp). The sense DNA strand coding for SNAP-25 RNAi-1 is TGAGTCCCTCGAGGCCGCCCGTCGCATGCTGC and GCAGCATGCTGC and GCAGCAT-GGACGCGGCGGCCTCGAGCGATCAGCC.

Data are presented as mean ± SEM.

Western blotting for p-SNAP-25 and SNAP-25 in HEK-293 cells and cortical neurons. For HEK-293 cells, 5 × 10^4 cells (ATCC) were seeded in 60 mm dish and transfected with WT or mutant SNAP-25/pIRE2-EGFP constructs (6 μg) using Lipofectamine 2000 (Invitrogen). Twenty-four hours after transfection, cells were treated with vehicle (DMSO control) or TPA (100 nM) for 15 min, scraped to one side, and pelleted by centrifugation. For neurons, one cortical hemisphere was trypsinized, triturated, and plated on to a 60 mm dish in Minimal Essential Medium (MEM, Invitrogen) for 24 h and switched to Neurobasal medium supplemented with B27 and GlutMax (Invitrogen). Neurons at 9 days in vitro (DIV) in culture were stimulated with TPA (100 nM) with or without bisindolylmaleimide I (Bis, 1 μM, Calbiochem), and with or without phosphatase inhibitors okadaic acid (100 nM, Calbiochem) and cyclosporin A (CsA, 2 μM) at 37°C for 15 min. TPA, Bis, and CsA were dissolved in DMSO as 10,000× stock solutions. Following treatment, neurons were rinsed once with ice-cold PBS and scraped to one side (Nunc) in PBS supplemented with 1% protease inhibitor (Sigma) and 1% phosphatase inhibitor cocktail 1 (Sigma). Following centrifugation at 15,000 g at 4°C for 10 min, pellets cells were lysed in lysis buffer containing the following (in mM): 25 Tris-HCl, 100 NaCl, 5 EDTA, 5 EGTA, 1% Triton X-100, 1% protease inhibitor, and 1% phosphatase inhibitor 1 (Sigma), pH 7.4, at 4°C for 30 min. Insoluble material was spun down at 16,000 × g for 15 min and protein content in the supernatant was subjected to BCA protein assay. Total protein (30 μg) was loaded on to a 4–12% Bis-Tris acrylamide gel (Invitrogen) and transferred for 1 h (Novex system, Invitrogen) on to a PVDF membrane. Membranes were blocked in 5% nonfat dry milk in TBST (0.05% Tween 20) for 30 min at room temperature (RT), incubated with an anti-p-SNAP-25 antibody (1:500; raised against the peptide MEKADS-PO4−-NKTRI corresponding to residues 182–192 of SNAP-25 including p-Ser187 in the center) (Iwasaki et al., 2000), or anti-SNAP-25 antibody (directed against the N terminus, 1:3000–5000, BD Transduction Laboratories), and detected with anti-rabbit (1:5000) or anti-mouse horseradish peroxidase (1:5000, GE Healthcare) secondary antibodies and enhanced chemiluminescence. Band intensities were quantified with Scion Image v. 4.0.3.2 (Scion) or ImageJ (NIH).

Antonia Follenzi, Albert Einstein College of Medicine). 293T cells in 15 mm plates were cotransfected with the transfer vector and packaging plasmids pMDL, pR aunque, and pKSV-G using the calcium phosphate method as described previously (Follenzi and Naldini, 2002). Forty-eight hours later, the medium was collected, filtered through a 0.45 μm filter, and centrifuged at 70,000 × g for 3 h. The viral pellet was resuspended in 200 μl of sterile PBS. All viral preparations were used after dilution to a concentration of 1:100 with medium.

Electrophysiology of NMDARs expressed in Xenopus oocytes. Adult female Xenopus laevis frogs (Nasco) were anesthetized in 0.15% 3-amino-benzonic acid ethyl ester (Sigma) and their oocytes were isolated with collagenase B (0.2% w/v) and prepared as described (Kushner et al., 1988). Stage V and VI oocytes were microinjected with 4–8 ng of NR1100/NR2A (1:3 by weight) cRNA. Two days after injection, they were injected with either WT or mutant SNAP-25 cRNA (20 ng). Two electrode whole-cell voltage-clamp recording was performed 1 d after the injection of SNAP-25 cRNA in Mg2+−free, Ca2+ or Ba2+ (1 mM) Ringer solution with microelectrodes filled with 3 M KCl, 10 mM HEPES, pH 7.2, with GeneClamp 500 (Molecular Devices, Foster City, CA). Data were digitized with pCLAMP6 (Molecular Devices). Responses were elicited by bath application of 300 μM NMDA with 10 μM glycine at a holding potential of −60 mV. Endogenous PKC was activated by bath application of the phorbol ester 12-O-tetradecanoylphorbol 13-acetate (TPA, 100 nM; Sigma) for 10 min. PKC potentiation was expressed as the ratio of the steady-state currents after TPA treatment to those obtained before TPA application. All experiments were performed at room temperature (RT). Data are presented as mean ± SEM.
were stimulated with TPA (100 nm) with or without Bis (1 μM) for 15 min. Cells were fixed in 4% paraformaldehyde for 5–10 min at RT, blocked, and permeabilized in 10% normal goat serum, 0.2% bovine serum albumin, and 0.1% Triton X-100 in PBS (contained 10 mM phosphate, 138 mM NaCl and 2.7 mM KCl, pH 7.4). The p-Ser187-SNAP-25 antibody was a gift from M. Takahashi (Kitsato University, Japan). Rabbit (p-SNAP-25, 1:250) and mouse (SNAP-25, 1:500, BD Biosciences/PharMingen) primary antibodies were incubated together at 4°C overnight, and were labeled with secondary antibodies conjugated to either Alexa, Fluor 488, Cy2 or Cy3 fluorophores (Jackson Immunoresearch or Invitrogen). Coverslips were mounted with an antifade reagent, ProLong (Invitrogen). Stained cells were visualized and photographed with an Olympus AX70 microscope using a 60X PlanApo oil-immersion lens (NA 1.40) and digitized using MagnaFire software (v. 2.1).

Quantification of p-SNAP-25 intensity levels was performed for selecting the entire HEK cell, rather than the leading edge, because it was not technically possible to arbitrarily draw the boundaries under basal conditions (low p-SNAP-25 levels). Thus, values as reported for HEK-293 cells may represent an underestimate of the true levels of p-SNAP-25 at the leading edge. Quantification of p-SNAP-25 intensity levels in neurons was performed by selecting the dendrites.

Electrophysiology of NMDA-elicited currents in hippocampal neurons. Dissociated cultures of hippocampal neurons were prepared from embryonic day 18 rats according to the methods of Banker and Goslin (1988). Hippocampal neurons (DIV 7–14) were transfected with 4 μg of WT or SNAP-25(S187A) in pRS2-E3EGF vector using a modified calcium phosphate method (Jiang et al., 2004) for 1–2 h that involved dissolution of the calcium phosphate precipitate in slightly acidic medium (Neurobasal) that had been equilibrated with 8% CO2. Twenty-four hours later, NMDA-elicited whole-cell currents were measured by 5 s bath application of 100 μM NMDA and 10 μM glycine. Near steady-state currents were measured at the end of the brief perfusion of agonists (5 s). The external solution contained the following (in mM): 145 NaCl, 2.5 KCl, 10 HEPES, 10 glucose, 2 CaCl2, pH 7.4 and −10 mm sucrose to adjust osmolality to 310–320 mOsm. Recording pipettes were pulled from borosilicate glass (A-M System) and open-tip resistance after heat polishing (Narishige) was typically 2–5 MΩ with an internal solution of (in mM): 120 CaCl2, 10 CsCl, 10 HEPES, 10 EGTA, 4 MgATP, 0.4 NaGTP, 0.1 CaCl2, 0.5 MgCl2, 5 phosphocreatine, adjusted to pH 7.3 with CsOH and 290–300 mOsm. The constitutively active form of PKC (PKM, 2 μM; Sigma) was dialyzed into the cell via its inclusion in the pipette, and NMDA currents were continually monitored and recorded using an Axopatch-1D amplifier (Molecular Devices) and digitized by the 1322A Digidata D/A converter (Axon) and pCLAMP 9.2 software. Signals were low-pass filtered at 2 kHz and digitized at 5 kHz. Vhold = −60 mV. Potentiation of NMDA currents was measured at 5 min after break-in and delivery of PKM and was expressed as a ratio to the amplitude of the basal current recorded at 0 min. SNAP-25 C-terminal peptides (70% purity) were custom-synthesized by Anaspec, and used at 10 μM in the pipette. Peptides were dissolved in water (1000X stocks), aliquoted and thawed for use on the day of experiment. The sequence for the blocking peptide was MEKADANKTRI and scrambled peptide was a combination of MEKADANKTRI and glycine, S-AMPA (25 μM; Sigma) was bath-applied for 1 st o simul-lation. For recording AMPAR- and NMDAR-mediated currents in the same cell, Cherylethrine (1 μM), an inhibitor of the catalytic domain of PKC, was applied at the end of experiments to confirm PKC activity. Clostridium botulinum neurotoxins (BoNT; from List Biological Laboratories) were reconstituted according to manufacturer’s instructions and applied to cells at 1 h at 37°C at 1 μg/ml (6.7 m) final concentration.

Electrophysiology of isolated NMDA EPSCs in hippocampus. Transverse hippocampal slices were prepared from P16-P21 C57BL/6 mice, as described previously (Christensen et al., 2004). Electrophysiological recordings were performed from neurons visually identified by infrared differential interference contrast microscopy using a 40X water-immersion objective. All experiments were performed at room temperature (22–25°C). Slices were continuously perfused with (in mM): 124 NaCl, 3 KCl, 1.25 KH2PO4, 1 MgCl2, 2 CaCl2, 26 NaHCO3, and 10 glucose, pH 7.3 (300 mOsM), supplemented with picrotoxin (50 μM), CNQX (10 μM) and LY303070 (the active isomer of GYKI-53655; 25 μM) to isolate synaptic responses mediated by NMDARs. Drugs were applied by switching between gravity-fed perfusion lines. EPSCs were evoked by paired-pulse stimulation (interstimulus interval of 100 ms) of mossy fibers by placing a bipolar electrode in the dentate granule cell layer. Eight-seal (>1 GΩ) whole-cell recordings were obtained from CA3 pyramidal neurons using patch pipettes pulled from borosilicate glass that had a resistance of 3–5 MΩ when filled with the following internal solution (in mM): 130 CaMgSO4, 4 NaCl, 10 HEPES, 0.2 EGTA, and 10 TEA, 5 QX-314, 4 ATP and 0.5 GTP (pH 7.3, 287 mOsM). PKC activators, PDBu (1 μM) or PKM (1 μM), and anti-SNAP25 antibody (20 mg/ml; SM181, Sternerberger Monoclonals), denatured (boiled) anti-SNAP25 antibody, SNAP-25 scrambled or blocking peptide (50 μM) alone or in combination were used in the patch pipette. Series resistance (<20 MΩ) was regularly measured during recordings, and cells were rejected if resistance changed >15% during the experiment. Data were filtered at 2 kHz, digitized, acquired through Axopatch 200A amplifier (Molecular Devices) and pClamp 8.0 software. Statistical significance was evaluated by the two-tailed unpaired Student’s t-test.

Western blot analysis of NMDAR surface expression. Cortical neurons in culture (DIV 9–11) were treated with TPA with or without phos-photase inhibitors, washed, and treated with membrane-impermeable EZ-Link Sulfo-NHS-SS-Biotin (Sulfosuccinimidyl 2-(biotinamido)-ethyl-1, 3-dithiopropionate, 1 mg/ml; Pierce) for 30 min at 4°C. Proteins were extracted in lysis buffer containing Tris-HCl (pH 7.40, 25 mM), NaCl (150 mM), EDTA (pH 8.0, 1 mM), SDS (0.1%), Na deoxycholate (0.5%), and protease and phosphatase inhibitors for 30 min at 4°C, and surface proteins were pulled down with NeutrAvidin beads (Pierce) ON/OFF at 4°C and released from the beads by boiling with DTT (1 mM). Total and surface proteins were subjected to SDS-PAGE and Western blotting using a monoclonal antibody 54.1 directed against the extracellular loop of the NR1 subunit (1:5000: gift from J.H. Morrison, Mount Sinai) or GluR1 (1:1000, Millipore AB1504).

Results

Ser187, but not Ser28 and Thr29, of SNAP-25 is an important PKC target

To examine a possible role for SNAP-25 as a PKC target critical to NMDAR insertion, we expressed NR1-4b/NR2A receptors together with WT and mutant SNAP-25 in Xenopus oocytes and recorded NMDA-elicited whole-cell currents before and after PKC activation. Xenopus oocytes are geometrically simple cells and provide a homogeneous source of NMDARs. NR1-4b/NR2A receptors exhibit the highest surface expression (Okabe et al., 1999) and highest PKC potentiation in oocytes (Durand et al., 1993). Application of the PKC-activating phorbol ester TPA (100 nm, 10 min) potentiated NMDA currents by ~8-fold in cells expressing NMDARs in the absence of exogenous SNAP-25 (control; Fig. 1a). PKC potentiation, defined as the ratio of steady-state NMDA current after PKC activation to that before PKC activation, was 8.30 ± 0.67 (n = 26, p < 0.001 vs control; in 1 mM Ca2+). The specific PKC inhibitor Ro-31-8220 (5 μM, 15 min preincubation) completely abolished TPA-induced potentiation of NMDA-elicited currents (from 4.30 ± 0.56 to 1.01 ± 0.29-fold, n = 3, p < 0.01; 1 mM Ba2+), indicating that TPA potentiation is PKC-mediated. Expression of wild-type (WT) SNAP-25 did not significantly alter basal or potentiated NMDA currents (Fig. 1b). Expression of dominant-negative SNAP-25 (lacking the last 20 amino acids in the C terminus, SNAP-25ΔD20 or dSNAP-25) (Yao et al., 1999; Lan et al., 2001a) markedly reduced basal NMDA currents (defined as the steadystate phase of the current before PKC potentiation and normalized to WT SNAP-25 current amplitudes; 25.7 ± 6.9% of WT, n = 20, p < 0.001; Fig. 1c) and PKC potentiation (normalized to WT SNAP-25 potentia-
and WT or mutant SNAP-25, 24 h after injection of SNAP-25 cRNA. NMDA currents were elicited by bath application of NMDA (N, 300 μM with 10 μM glycine; gray bars above current traces) and recorded by two-electrode voltage clamp at Vh = −60 mV in 1 mM Ca^{2+} and 0 Mg^{2+}.

Application of the PKC-activating phorbol ester TPA (100 nM, 10 min, black bar) potentiated NMDA currents by −8-fold in control (H2O-injected) oocytes. PKC potentiation is defined as the ratio of steady-state NMDA current after to that before PKC activation.

PKC phosphorylation, a role for PKC-dependent phosphorylation of SNAP-25 in NMDAR potentiation is unclear. To examine the possibility that SNAP-25 is a functionally relevant target of PKC phosphorylation and to identify residue(s) in SNAP-25 that are critical to PKC potentiation of NMDARs, we expressed NMDARs together with WT or mutant SNAP-25 lacking one or more of its putative phosphorylation sites (Ser187, Ser28, Thr29) in *Xenopus* oocytes. Replacement of the serine residue 187 in SNAP-25 with an alanine (Ser187Ala or S187A) markedly reduced basal NMDA currents (27.5 ± 3.4% of WT, n = 22, p < 0.001; Fig. 1d) and degree of PKC potentiation (59.9 ± 7.5% of WT, n = 22, p < 0.01; Fig. 1d), reductions similar to those observed for dnSNAP-25Δ20. Thus, in oocytes, a single point mutation in SNAP-25 is sufficient to suppress constitutive and regulated NMDAR exocytosis. In contrast, elimination of the other two putative sites of PKC phosphorylation, Ser28 and Thr29, even in combination did not detectably alter either basal or potentiated currents (basal currents, 119 ± 9% of WT, n = 10, p > 0.05; PKC potentiation, 94.5 ± 4.8% of WT, n = 10, p > 0.05; Fig. 1e). These findings indicate that Ser187 is the exclusive site of PKC phosphorylation in SNAP-25 that is functionally relevant to NMDAR insertion. The phosphomimetic mutant SNAP-25(S187D), which mimics phosphorylation by replacement of Ser with the negatively charged residue aspartate (D), markedly enhanced basal NMDA currents even in the absence of PKC activation (186 ± 31% of WT, n = 5, p < 0.05; Fig. 1f), but did not detectably alter PKC potentiation (88.0 ± 6.1% of WT, n = 5, p > 0.05; Fig. 1f–i). The fact that mutation of Ser187 into Asp, a non-phosphorylatable residue, does not affect PKC potentiation suggests that Ser187 of SNAP-25 may not be the only target of PKC phosphorylation involved in NMDAR trafficking. These results suggest that Ser187 of SNAP-25 is a target of PKC phosphorylation relevant to NMDAR potentiation in oocytes.

PKC phosphorylates SNAP-25 at Ser187

To directly show that PKC phosphorylates SNAP-25 at Ser187, we performed two experiments. First, we expressed WT and mutant SNAP-25(S187A) under the pRES2-EGFP vector, which allows transfected cells to be identified by means of its green fluorescence, in HEK-293 cells and assessed PKC-induced phosphorylation by Western blot analysis. Application of PKC-activating phorbol ester TPA (100 nM, 15 min) to HEK-293 cells induced phosphorylation of SNAP-25 at Ser187, as assessed by Western blots probed with a phospho-specific antibody (Fig. 2a,

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**Figure 1.** Ser187, but not Ser28 and Thr29, of SNAP-25 is critical to PKC potentiation of NMDA currents. a–f, Representative experiments showing PKC potentiation of NMDA-elicited currents recorded from *Xenopus laevis* oocytes expressing NR1-4b/NR2A and WT or mutant SNAP-25, 24 h after injection of SNAP-25 cRNA. NMDA currents were elicited by bath application of NMDA (N, 300 μM with 10 μM glycine; gray bars above current traces) and recorded by two-electrode voltage clamp at Vh = −60 mV in 1 mM Ca^{2+} and 0 Mg^{2+}.

a, Application of the PKC-activating phorbol ester 12-O-tetradecanoylphorbol 13-acetate (TPA, 100 nM, 10 min, black bar) potentiated NMDA currents by −8-fold in control (H2O-injected) oocytes. PKC potentiation is defined as the ratio of steady-state NMDA current after to that before PKC activation. b, Expression of WT SNAP-25 does not detectably alter PKC potentiation of NMDA-elicited currents. c, The dominant-negative mutant of SNAP-25, SNAP25Δ20, markedly reduced the basal NMDA current and PKC potentiation. Note the change in scale. d, A point mutation in which Ser187 was replaced by Ala (S187A) in SNAP-25 reduced both basal NMDA currents and PKC potentiation to an extent indistinguishable from SNAP-25Δ20. e, Replacement of Ser28 and Thr29 by Ala (SNAP-25 S28A/T29A) did not detectably alter basal NMDA currents or PKC potentiation. f, The phosphomimetic mutant, SNAP-25 S187D, markedly enhanced basal NMDA currents without affecting PKC potentiation. g, h, Summary bar graphs showing effects of WT and mutant SNAP-25 on basal NMDA currents (g) and PKC potentiation (h). PKC potentiation in un.injected oocytes and oocytes expressing mutant SNAP-25 constructs was normalized to that in cells expressing WT SNAP-25. Numbers on bars indicate the number of oocytes (n) involving 3–5 independent experiments performed from different batches of oocytes. Data represent means ± SEM. *p < 0.05; **p < 0.01; ***p < 0.001.

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top). Coapplication of the potent and specific PKC inhibitor bis-indoylmaleimide I (Bis, 1 μM) completely abolished TPA-induced phosphorylation of SNAP-25, confirming that TPA acts via PKC to exert its effects (Fig. 2a). In contrast, cells expressing mutant SNAP-25(S187A) exhibited robust SNAP-25 expression, but little or no p-SNAP-25 either before or after application of TPA (Fig. 2a). Stripping and reprobing the membrane with an antibody directed against full-length SNAP-25 revealed equal loading of samples and absence of SNAP-25 in untransfected cells, indicating the specificity of the antibody.

Second, we expressed WT and mutant SNAP-25(S187A) in HEK-293 cells and performed immunolabeling of total and p-SNAP-25. Under basal conditions, SNAP-25 localized to the plasma membrane, as well as to the Golgi apparatus (perinuclear labeling); little or no p-SNAP-25 was detected (Fig. 2b). Application of TPA did not detectably alter the abundance or cellular localization of total SNAP-25, but produced a dramatic increase in p-SNAP-25 immunofluorescence, prominent at the edges of the cells (Fig. 2c). The specific PKC inhibitor Bis again abolished TPA-induced phosphorylation of SNAP-25, indicating that SNAP-25 phosphorylation is PKC-mediated (Fig. 2d). In contrast, cells expressing mutant SNAP-25(S187A) exhibited robust SNAP-25 immunofluorescence, but little or no p-SNAP-25 immunofluorescence either before or after application of TPA (Fig. 2e–g). These findings demonstrate that PKC promotes phosphorylation of SNAP-25 at Ser187 in HEK-293 cells. Although we cannot rule out additional sites of phosphorylation at Ser28 and Thr29, data in Figure 1 indicate that phosphorylation of these sites to occur, it would not be functionally relevant to PKC potentiation.

The results thus far demonstrate that SNAP-25 is a functionally relevant target of PKC involved in PKC potentiation of NMDARs in oocytes and that PKC phosphorylates SNAP-25 at Ser187 in HEK-293 cells. To examine whether Ser187 of SNAP-25 is a target of PKC phosphorylation in neurons, we performed two experiments. First, we treated cortical neurons (DIV 19) with TPA or vehicle and subjected cell lysates to Western blot analysis. Cortical neurons exhibited little or no p-SNAP-25 basally, whereas upon TPA stimulation, p-SNAP-25 was markedly induced (Fig. 3a). These findings are in confirmation of observations in PC12 cells (Iwasaki et al., 2000), chromaffin cells (Nagy et al., 2002) and HEK-293 cells (Fig. 2). Application of okadaic acid (OA, 100 nM) with cyclospermine A (CsA, 2 μM), a mixture that inhibits protein phosphatases 1 (PP1), PP2A, and PP2B (also known as calcineurin), enhanced TPA-elicted phosphorylation of SNAP-25 at Ser187 (Fig. 3a, top); the mixture alone did not induce phosphorylation of SNAP-25 at Ser187 (data not shown). Application of the specific PKC inhibitor Bis completely abolished TPA-induced phosphorylation of SNAP-25 at Ser187 in the absence or presence of phosphatase inhibitors (Fig. 3a, top). Summary data indicate that the phosphatase inhibitors enhanced TPA-induced phosphorylation of SNAP-25 by ~2-fold and that Bis completely abolished phosphorylation (TPA vs control, TPA + Bis vs TPA and TPA + CsA + OA + Bis vs TPA, n = 4, p < 0.001; TPA + CsA + OA vs TPA, n = 4, p < 0.05; Fig. 3b). In contrast, neither PKA inhibitors (PKI(14-22) and H-89) nor a CaMKII inhibitor (KN-93) attenuated SNAP-25 phosphorylation in neurons (supplemental Fig. 1, available at www.jneurosci.org as supplemental material). These findings strongly suggest that the TPA-induced phosphorylation of SNAP-25 is PKC-dependent in HEK-293 and neurons.

Second, we performed immunolabeling of hippocampal neurons (DIV 14) with antibodies to SNAP-25 and p-SNAP-25. Under basal conditions, SNAP-25 immunolabeling was prominent in cell bodies and processes of neurons (Fig. 3c,d). In contrast, p-SNAP-25 immunolabeling was relatively low, presumably due to low basal PKC activity and/or high phosphatase activity. Application of TPA did not detectably alter the abundance or cellular localization of total SNAP-25, but markedly increased pSNAP-25 immunofluorescence (Fig. 3c,d). Quantification of fluorescence intensity showed that TPA increased the p-SNAP-25/total SNAP-25 ratio in dendrites by ~50% (control vs TPA; p-SNAP-25: 100 ± 11% vs 150 ± 9%, p < 0.01; total SNAP-25: 100 ± 17% vs 81 ± 15%, p > 0.4; p-SNAP-25/total SNAP-25: 100 ± 10%, vs 229 ± 40%, n = 7, p < 0.05; Fig. 3d). These findings indicate that PKC phosphorylates SNAP-25 at Ser187 in neurons and that after PKC stimulation, p-SNAP-25 is enhanced in neuronal processes. The results do not exclude the possibility that PKC also phosphorylates other targets such as the
PKC-mediated insertion of NMDARs in neurons is SNAPRE-dependent

The results reported thus far demonstrate that PKC phosphorylates SNAP-25 at Ser187 in neurons, but do not implicate SNAP-25 as a functionally relevant substrate of PKC phosphorylation in neurons. Toward this end, we undertook three experimental approaches. First, we monitored PKC potentiation of NMDA-elicited currents by whole-cell patch-clamp recording of neurons with BoNT A (200 ng/ml; pretreatment for 1 h) which cleaves v-SNARE VAMP2/synaptobrevin-2, also blocked PKM potentiation of NMDA currents (inactive vs active BoNT B: peak, 131 ± 16% vs 99 ± 4%; n = 9; p < 0.001; steady state, 140 ± 14% vs 106 ± 5%, n = 9; p < 0.02 vs basal currents; Fig. 4a–c), presumably due in large part to enhanced delivery of NMDARs to the cell surface (Lan et al., 2001a). Treatment of neurons with BoNT B (200 ng/ml; pretreatment for 1 h) did not detectably alter basal NMDA currents (data not illustrated), but abolished PKC potentiation of NMDA currents of both peak and steady-state (SS) phases of NMDA currents in hippocampal neurons (inactive vs active BoNT A: peak, 157 ± 16% vs 99 ± 2%, n = 9; p < 0.001; steady state, 140 ± 14% vs 106 ± 5%, n = 9; p < 0.02; Fig. 4a–c). Consistent with SNAPRE-dependent receptor exocytosis, pretreatment with BoNT B (200 ng/ml), which cleaves v-SNARE VAMP2/synaptobrevin-2, also blocked PKM potentiation of NMDA currents (inactive vs active BoNT B: peak, 131 ± 5% vs 97 ± 14%, n = 4; p < 0.05; supplemental Fig. 2, available at www.jneurosci.org as supplemental material). These findings indicate that PKC potentiation of NMDA responses in hippocampal neurons occurs via SNAP-25-mediated, SNAPRE-dependent exocytosis.

RNAi-mediated depletion of SNAP-25 blocks PKC potentiation of NMDARs

Second, we examined with the impact of RNAi-mediated knockdown of SNAP-25 on PKC potentiation of NMDA currents. We
designed two RNAi sequences directed against mouse SNAP-25 cDNA and evaluated them separately for their ability to knockdown SNAP-25 expression in HEK-293 cells. A generic RNAi that does not target any known vertebrate gene (non-targeting (NT)-RNAi), served as a negative control (pcDNA6.2). Both SNAP-25 RNAi-1 (directed to bps 470–490 of mouse SNAP-25) and SNAP-25 RNAi-2 (directed to bps 681–701 of SNAP-25), but not NT RNAi, markedly attenuated recombinant SNAP-25 protein expression in HEK-293 cells (48 h after transfection; RNAi-1: to 17% of control; RNAi-2: to 30% of control; 72 h after transfection: both to 4% of control; supplemental Fig. 3, available at www.jneurosci.org as supplemental material). To validate the specificity and effectiveness of the SNAP-25 RNAi-2 sequence in cells with neuronal traits, we examined its impact on endogenous SNAP-25 and SNAP-23 abundance in Neuro2A (N2A) cells. N2A cells were transfected with a lentiviral transfer vector driving cocistronic expression of EGFP and SNAP-25 RNAi-2 (hereafter referred to as “SNAP-25 RNAi”). Seventy-two hours after transfection, cells were lysed and subjected to Western blotting for SNAP-25. SNAP-25 RNAi was highly effective in that it markedly reduced SNAP-25 protein abundance relative to that of untransfected cells (not illustrated) or cells transfected with NT (control) RNAi (to 18%), and was specific in that it did not detectably alter SNAP-23 abundance (Fig. 4d).

To examine the effects of SNAP-25 RNAi on NMDA currents in a physiological milieu, we next infected neurons with lentivirus containing SNAP-25 RNAi or NT RNAi (control) and monitored PKM-induced potentiation of NMDA currents. At 48–72 h after transduction, SNAP-25 RNAi was robustly expressed, as evidenced by intense EGFP fluorescence (data not illustrated). SNAP-25 RNAi induced little effect on basal NMDA currents, but markedly reduced PKM potentiation of both peak and steady-state phases of NMDA currents neurons relative to WT SNAP-25. A sequence directed against SNAP-25, but not an unrelated sequence (control), markedly reduced SNAP-25 expression. Striping and reprobing for β-actin revealed equal loading and that SNAP-23 expression is unaffected. *Nonspecific band.

Phosphorylation of SNAP-25 at Ser187 is critical to PKC insertion of NMDARs in neurons

Third, we expressed WT or mutant SNAP-25(S187A) via the pIRE52-EGFP vector in cultured hippocampal neurons and monitored PKC potentiation of NMDA-elicted currents. The results thus far implicate a role for SNAP-25 in NMDAR insertion in neurons, but do not address the target of PKC phosphorylation. In neurons expressing WT SNAP-25, PKM potentiated NMDA currents by ~80% relative to basal currents within 5–10 min of break-in (potentiation of the peak to 186 ± 33%; p < 0.001; potentiation of steady state to 199 ± 16%; p < 0.001; Fig. 5a,b,d). Expression of SNAP-25(S187A) produced no significant change in basal NMDA-elicted currents relative to WT SNAP-25 (WT vs S187A, peak, 926 ± 211 vs 635 ± 254 pA, n =
Phosphorylation of postsynaptic SNAP-25 at Ser187 is critical to PKC-induced insertion of NMDARs in neurons. A, Representative traces showing PKC potentiation of NMDA responses in neurons expressing SNAP-25/pIRES2-EGFP. Whole-cell patch-clamp recordings were performed on hippocampal neurons (DIV 7–16) expressing SNAP-25 WT or S187A/pIRES2-EGFP, 1 d after transfection. PKM in the patch pipette potentiated NMDA currents expressing SNAP-25 WT (left). Mutant SNAP-25 (S187A) abolished PKC potentiation of NMDA currents (right). B, Summary time course of PKM potentiation in cells expressing WT (●) versus mutant SNAP-25 (S187A) (○). PKM potentiated the peak (top) and steady-state (SS; bottom) phases of NMDA-elicited currents (n = 5 each). Current amplitudes were normalized to amplitudes of control responses recorded just after break-in (at 0 min). C, D, Expression of SNAP-25(S187A) did not alter the peak or steady-state phase of basal NMDA currents (C), but abolished PKM-induced potentiation (D). n.s., not significant; *p < 0.05; **p < 0.01.

To examine whether PKC-induced phosphorylation of SNAP-25 selectively regulates insertion of NMDARs or more generally regulates delivery of ligand-gated ion channels to the cell surface, we monitored the impact of PKM on α-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid receptor (AMPA)-mediated currents in the same cell. After washout of NMDA and glycine, we recorded currents elicited by 1 s bath application of S-AMPA (25 μM) to hippocampal neurons. Whereas PKM potentiated NMDA-elicited currents in the presence of scrambled peptide (Fig. 6b,c), it produced no detectable effect on AMPA-elicited currents (blocking, 100.8 ± 3.3% of control; scrambled, 92.2 ± 3.8% of control; p > 0.05 for all; Fig. 6d–f right). These findings strongly suggest that postsynaptic assembly of the SNARE complex is required for PKC-dependent insertion of NMDARs (but not AMPARs) in hippocampal neurons.

SNAP-25 mediates PKC potentiation in hippocampal slices

The experiments thus far establish a role for SNAP-25 in PKC-dependent insertion of NMDARs, but do not distinguish between synaptic and extrasynaptic NMDARs. To address this issue, we recorded isolated NMDA EPSCs (EPSCs) at mossy fiber (mf)-CA3 synapses of acute hippocampal slices from juvenile rats in the presence of CNQX and LY303070 (the active isomer of GYKI-53655) to block kainate receptors and AMPARs, and bicuculline to block GABA\(_B\)Rs. The PKC-activating phorbol ester PDBu (1 μM), delivered via the recording pipette, did not detectably alter the paired-pulse ratio [PPR; control, PPR at 0 min, 2.17 ± 0.32; PPR at 5–10 min, 2.31 ± 0.28; scrambled, PPR at 0 min, 2.50 ± 0.39, Fig. 7a; scrambled, PPR at 5–10 min, 2.76 ± 0.48 (data not illustrated)], but markedly potentiated NMDA EPSCs in the absence (data not illustrated) or presence of the scrambled peptide (control, to 228.8 ± 29.9%, n = 9, p < 0.01; scrambled, to 235.4 ± 42.6%, n = 7, p < 0.01; Fig. 7b,d). As in the case of NMDA-elicited whole-cell currents recorded from dissociated neurons, application of the SNAP-25 blocking peptide via the patch pipette did not significantly affect basal NMDA transmission, assessed at 5 min after break-in (blocking peptide, 94.1 ± 15%; n.s. vs basal current in the absence of the peptide), but greatly reduced PDBu potentiation of the NMDA EPSC at mf-CA3 synapses (scrambled peptide, 235.4 ± 42.6%; blocking peptide, 105.7 ± 8.2%; n = 7, p < 0.05 vs scrambled peptide; Fig. 7b,d). Additionally, dialysis of PKM (1 μM) into the postsynaptic
cell potentiated NMDA EPSCs to 364 ± 76.5% of control by 5–10 min after break-in (Fig. 7c,d). Delivery of an anti-SNAP-25 antibody (α-SNAP-25) together with PKM greatly reduced the PKM-induced potentiation of NMDA EPSCs (from 364 ± 76.5%, n = 8, to 167.4 ± 18.9% n = 6, p < 0.05; Fig. 7c,d). These findings demonstrate a postsynaptic action of SNAP-25 in PKC-mediated potentiation of synaptic NMDARs at mf-CA3 synapses.

**Activation of PKC promotes NMDAR surface expression in neurons**

The results thus far indicate that PKC potentiates postsynaptic NMDARs at hippocampal synapses, but do not address the mechanism by which the potentiation occurs. To address this issue, we examined the impact of PKC activation on surface expression of NMDARs in neurons. Application of TPA in the presence of the phosphatase inhibitors OA (100 nM) and CsA (2 μM) (but not OA and CsA alone) enhanced the surface expression of NMDAR subunit NR1 in cortical neurons (DIV 7–10), as assessed by Western analysis of biotinylated cell-surface proteins (Fig. 8a,b). The increase in NR1 cell surface expression was blocked by Bis, indicating that it is PKC-mediated, and by BoNT A, indicating that it is SNARE-dependent [p < 0.05 for TPA+OA+CsA, n = 4 vs control (nontreated), n = 4; Fig. 8h,c]. Total NR1 levels were not detectably altered (Fig. 8h,c). In contrast, TPA in the presence of OA and CsA did not significantly enhance the surface expression of AMPARs as assessed by Western blotting for biotinylated surface GluR1 or GluR2 (GluR1: OA+CsA vs OA+CsA+TPA, 104 ± 16% vs 100 ± 9%, n = 3,
currents (NMDA EPSCs) were recorded from CA3 pyramidal neurons in the presence of CNQX, LY303070, and picrotoxin (to block 6–8 experiments in which responses were binned every minute.

SNARE-Dependent NMDAR Trafficking

Whereas SNAP-25 is critical to regulated, PKC-dependent exocytosis of NMDARs, SNAP-23 mediates constitutive exocytosis of NMDAR transport packets in cortical neurons prior to synapse formation (Washburn et al., 2004) and constitutive cycling of NMDARs at CA1 synapses of cultured hippocampal slices (Suh et al., 2009). SNAP-25 and syntaxin-1 (target or t-SNAREs) localize to the plasma membrane, where they dock with vesicular “v-SNAREs” such as VAMP2 and mediate fusion of cargo-loaded vesicles with the plasma membrane (Südhof and Rothman, 2009). PKC phosphorylation of SNAP-25 at Ser187 promotes assembly of the SNARE complex and thereby facilitates recruitment of vesicles at the presynaptic membrane (Nagy et al., 2002). Based on findings in the present study, we propose a model whereby PKC phosphorylation of SNAP-25 at Ser187 also promotes insertion of NMDA-EPSCs at this synapse (Kwon and Castillo, 2008; Rebola et al., 2008). These findings, together with our finding that SNAP-25 is a target of PKC phosphorylation functionally relevant to NMDAR trafficking, provide a molecular mechanism by which activity induces LTP of NMDA-EPSCs at this synapse (Kwon and Castillo, 2008; Rebola et al., 2008).

SNARE-dependent glutamate receptor trafficking

The specificity of PKC for SNAP-25 phosphorylation and NMDAR insertion is indicated by several findings. First, phosphorylation of Ser187, but not Ser28 and Thr29, is critical to PKC-dependent insertion of NMDARs in oocytes and neurons. Second, RNAi against SNAP-25 (but without effect on SNAP-23) abolishes PKC potentiation of NMDARs in neurons. Third, whereas PKC potentiates NR2A- and NR2B-containing NMDARs, it

Discussion

The present study shows that PKC promotes NMDAR insertion of new NMDA channels in hippocampal neurons in culture and at mf-CA3 synapses via SNARE-dependent exocytosis. Although the functional target of PKC potentiation is well established, the molecular target of PKC is unclear. We further show that SNAP-25, a member of the SNARE family of vesicular fusion proteins, is a target of PKC phosphorylation functionally relevant to NMDAR trafficking. Several independent findings indicate that PKC-induced phosphorylation of SNAP-25 at Ser187 (and assembly of the SNARE complex) are critical to regulated delivery of NMDARs at the cell surface. First, dominant negative SNAP-25 reduced both basal and, in corroboration of Lan et al. (2001a), potentiated NMDA-elicted currents in oocytes. Second, RNAi-mediated depletion of SNAP-25 and BoNT A-mediated cleavage of SNAP-25 abolished PKM potentiation of NMDA currents in neurons. Third, overexpression of SNAP-25(Ser187Ala) abolished PKC potentiation of NMDA-elicted currents in oocytes and neurons. Fourth, acute disruption of SNAP-25 function by introduction of a blocking peptide corresponding to the C terminus of SNAP-25 phosphorylated at Ser187 attenuated PKC-induced insertion of NMDARs in hippocampal neurons and slices. Fifth, introduction of a function-blocking SNAP-25 antibody into the postsynaptic cell markedly inhibited PKC potentiation of the NMDA EPSCs at CA3 synapses in acute hippocampal slices. Our findings that PKC promotes insertion of new NMDA channels at the postsynaptic side of mf-CA3 synapses in a SNARE-dependent manner are consistent with findings of others (Kwon and Castillo, 2008). These findings, together with our finding that SNAP-25 is a target of PKC phosphorylation functionally relevant to NMDAR trafficking, provide a molecular mechanism by which activity induces LTP of NMDA-EPSCs at this synapse (Kwon and Castillo, 2008; Rebola et al., 2008).
does not potentiate NR2C- or NR2D-containing receptors in *Xenopus* oocytes (Mori et al., 1993). Fourth, PKC potentiation is greater for NMDARs containing the short NR1 splice variants, NR1-3 and NR1-4, versus NMDARs containing long NR1 isoforms (Durand et al., 1992, 1993). Fifth, whereas PKM promotes postsynaptic SNAP-25-dependent insertion of NMDARs (present study) and SNAP-25-dependent internalization of kainate receptors (Selak et al., 2009), it does not alter AMPAR trafficking (present study). These findings are consistent with findings that high-frequency stimulation (HFS) potentiates NMDA, but not AMPA, synaptic currents in a PKC-dependent manner at mf-CA3 synapses (Kwon and Castillo, 2008). Whereas conventional PKM potentiates NMDA but not AMPA currents, atypical PKMzeta promotes surface expression of AMPARs but not NMDARs (Ling et al., 2006). The catalytic site of PKMzeta differs structurally from that of conventional and novel PKCs and would thus be expected to affect distinct molecular targets (Sacktor, personal communication). Our findings are also consistent with findings that NMDARs and AMPARs are transported to synaptic sites via distinct vesicles (Fong et al., 2002; Washbourne et al., 2002b) and are sorted via different intracellular pathways (Jeyifous et al., 2009) in hippocampal neurons. Whereas AMPARs undergo forward trafficking to the plasma membrane via the conventional somatic Golgi network, NMDARs traffic via a nonconventional secretory pathway involving CASK, SAP97 and dendritic Golgi outposts (Jeyifous et al., 2009) in close proximity to NMDARs and far from intracellular AMPARs.

It is well established that PKC acts via Src to regulate NMDAR function (Lu et al., 1999), and that genetic deletion of Src blocks PKM potentiation (Lu et al., 1999). PKC phosphorylates and activates Src (Lu et al., 1999), which in turn phosphorylates Tyr residues on the C-terminal tails of NR2 subunits, thereby increasing channel open probability and mean open time (Yu et al., 1997). Although Fyn kinase-dependent phosphorylation of NR2B at Tyr1472 promotes constitutive delivery of synaptic NMDARs (Prybylowski et al., 2005), Src itself does not regulate NMDAR trafficking. A possible scenario is that whereas PKC acts via SNAP-25 to regulate NMDA trafficking (Lan et al., 2001a), it acts via Src to regulate intrinsic NMDA channel function. In that event, loss of either SNAP-25 or Src would be sufficient to attenuate PKC potentiation of NMDA currents.

We found that the phosphomimetic mutant SNAP-25(S187D) did not detectably alter basal NMDA currents or PKC-mediated trafficking of NMDARs in neurons. Although phosphomimetic mutants may mimic the actions of phosphorylated proteins, in many cases they do not (Richard L. Huganir, Leonard K. Kaczmarek, personal communications). Given that an aspartate (or glutamate) residue approximates the negative charge, but is not structurally identical to, a phosphorylated serine, this is not surprising. Thus, our negative findings in the case of SNAP-25(S187D) argue neither for nor against a role of SNAP-25 S187 phosphorylation in PKC potentiation of NMDA currents in neurons. Our finding that neither the (S187A) and (S187D) mutants affects basal NMDA currents in neurons is consistent with findings that the SNAP-25 blocking peptide does not alter basal synaptic NMDA currents at CA3 synapses (present study; Kwon and Castillo, 2008) and that overexpression of the SNAP25(S187A) mutant does not detectably alter basal NMDA currents. Our finding in neurons is in contrast to our finding that the S187D mutant potentiates NMDA currents and partially occludes PKC potentiation in oocytes. There are several possible explanations for the differences in oocytes versus neurons. First, whereas the oocytes express a pure population of NR1/NR2A receptors, neurons express heterogeneous NMDARs. Second, synaptic receptors localize to postsynaptic densities together with scaffolding, adaptor and signaling proteins which modify their trafficking and function and are not necessarily expressed in oocytes. Third, whereas we bath-applied phorbol esters to oocytes, we delivered recombinant PKM via the patch pipette to neurons. Fourth, neurons may employ different mechanisms for constitutive versus regulated trafficking of NMDAs. The experiments in neurons are more physiologically relevant and more likely to reflect PKC-dependent regulation of NMDARs and

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**Figure 8.** PKC activation enhances surface expression of NMDARs via SNAP-25.  
**a**, Timeline of experiment. After 45 min incubation with BoNT A (200 ng/ml), cortical neurons (DIV 7–10) were treated with phospactase inhibitors OA (100 nm) and CsA (2 μm), followed by TPA to activate PKC. Fifteen minutes after TPA incubation, cell surface proteins were biotinylated with EZ-Link Sulfo NHS-SS, cells lysed and surface proteins pulled down with NeutrAvidin beads and subjected to Western blotting for NR1. **b**, TPA-induced insertion of NMDARs at the cell surface is PKC-mediated and SNAP-25-dependent. Representative Western blot showing that TPA + OA + CsA enhanced surface expression of NMDARs relative to control (untreated) or OA and CsA alone; the increase in surface expression was blocked by Bis and BoNT A. Treatments did not alter total NR1 levels. **c**, Summary bar graph showing that BoNT A significantly inhibited the increase in NMDAR surface expression induced by TPA + OA + CsA (n = 4), *p* < 0.05. **d**, Representative Western blots probed for GluR1 and GluR2 (d) and summary bar graph (e) showing that TPA, in the presence of OA and CsA, does not enhance surface expression of AMPARs. **f**, Proposed model showing that activation of PKC phosphorylates SNAP-25 and promotes insertion of new NMDA, but not AMPA, channels at the cell surface, consistent with trafficking of AMPARs and NMDARs in distinct postsynaptic vesicles and delivery to the plasma membrane via distinct pathways of exocytosis.
Regulated NMDAR trafficking in synaptic plasticity

Recent studies indicate that synaptic NMDAR number and/or subunit composition are also regulated in response to neuronal activity and sensory experience (Wenthold et al., 2003; Pérez-Otaño and Ehlers, 2005; Lau and Zukin, 2007), and that NMDARs serve not only as the trigger of synaptic plasticity, but may also contribute to the expression of LTP and LTD (Grosshans et al., 2002; Watt et al., 2004). In adult rats, HFS promotes PKC-dependent, SNARE-dependent insertion of NMDARs at mGluR4 synapses (LTP_NMDAR). (Kwon and Castillo, 2008). Given that LTP-inducing protocols can induce SNAP-25 phosphorylation (Genoud et al., 1999), HFS may act via phosphorylation of SNAP-25 to promote insertion of NMDARs and elicit LTP_NMDAR. In addition to activation of NMDARs, stimulation of group I mGluRs promotes delivery of NMDA channels to the cell surface (Lan et al., 2001b). These observations raise the possibility that mGluRs could induce PKC-dependent phosphorylation of SNAP-25 and promote NMDAR insertion at the plasma membrane. Regulation of NMDAR trafficking provides a powerful mechanism for altering transmission efficacy and neuronal excitability during synaptic plasticity and remodeling.

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**Supplementary Fig. 1.** TPA-induced phosphorylation of SNAP-25 is mediated by PKC. Cortical neurons in culture were treated with TPA (100 nM, 15 min) in the absence or presence of various kinase inhibitors. Cell lysates were subjected to Western blotting and probed with p-SNAP-25 (upper panel). TPA induced phosphorylation of SNAP-25, which was completely abolished when pre-treated with (15 min) the specific PKC inhibitor bisindoylmaleimide I Bis (1 µM) and markedly inhibited by another PKC inhibitor Chelerythrine (Chel, 10 µM). PKA inhibitors (PKI14-22, PKI, 1 µM and H89, 10 µM) and a CaMKII inhibitor (KN93, 10 µM) did not markedly affect SNAP-25 phosphorylation. Stripping and reprobing with an antibody against total SNAP-25 reveals equal loading of samples (lower).

**Supplementary Fig. 2.** VAMP2/synaptobrevin-2 is required for PKM potentiation. (a) Hippocampal neurons were pretreated with BoNT B (200 ng/ml, 1 h), which specifically cleaves VAMP2, and NMDA currents were monitored in the presence of PKM in the patch pipette. Active (○), but not inactive (■), BoNT B abolished PKM potentiation of both peak and steady-state (SS) phases of NMDA currents in hippocampal neurons (b,c) Summary time-course (b) and bar graph (c) of data shown in a. n = 4; *, P < 0.05; n.s., non-significant.

**Supplementary Fig. 3.** RNAi-mediated knockdown of recombinant SNAP-25. Two DNA sequences directed against independent regions of SNAP-25 were subcloned into the pcDNA6.2-GW/EmGFP vector (pcDNA6.2). HEK-293 cells were mock-transfected (no SNAP-25 or pcDNA6.2), co-transfected with pcDNA6.2 and SNAP-25 or pcDNA6.2 harboring 2 different RNAis that target different sequences of the SNAP-25 cDNA. Both SNAP-25 RNAi sequences induced robust knockdown of SNAP-25 expression at 48 and 72 h, as assessed by Western blotting. Stripping and reprobing with an antibody against β-actin reveals equal loading of samples in all lanes and the presence of GFP in cells expressing pcDNA6.2 constructs.

**Supplementary Fig. 4.** Normal PKM-induced potentiation of NMDA currents in neurons expressing SNAP-25 S187D. We recorded NMDA-evoked current in hippocampal neurons (DIV12-14) expressing SNAP-25 S187D/pIRES2-EGFP in the presence of PKM in the patch pipette. Expression of mutant SNAP-25 S187D allowed PKM-induced potentiation of NMDA currents.
Supplementary Figure-1 Zukin
Supplementary Figure-3 Zukin
SNAP-25 S187D

1 min 4 min
N N

500 pA
5 s

Normalized current

Peak
Steady-state
PKM

Supplementary Figure-4 Zukin